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# Exposed Proteomes of Brachyspira pilosicoli responsible for the overall decline in poultry and pig's meat *production*

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# **ABSTRACT**

Brachyspira pilosicoli is a gram-negative spiral-shaped, obligate anaerobe bacterium belongs to the spirochete family, the pathogen causing diarrhea, reduced growth rate, reduced egg and meat production, and quality. In recent years, there has been a substantial rise in the rate of Brachyspira pilosicoli infections, as well as the emergence of the virulent and antibiotic-resistant Brachyspira pilosicoli strains. So there is an urgent need for the identification of the therapeutic potential target and development of new drugs for the treatment and prevention of Brachyspira Pilosicoli infections. In the current investigation, we used approach by reference protein (RefSeq. Protein) the individual 1378 protein sequence was entered in DELTA-BLAST against proteomes of pigs (taxid: 9821) pigs & lt; 2 & gt; (taxid: 9823), chickens (taxid: 9031) and result of 27 protein sequences no similarity was found, these 27 protein sequences were again compared against the database of essential single genes individually and 9 of them had no similarity and 18 protein sequences were taken out which had similarity and their protein name, biological process, molecular function and role in virulence have been mentioned and discussed.

**Keywords**— Brachyspira, Zoonotic

## 1. INTRODUCTION

The seven officially named and several unofficially named species of the anaerobic *spirochetes* are colonizing the large intestine of the mammals and birds. These seven official species belong to the genus *Brachyspira* [Stanton TB et al 2006]. Here *Brachyspira hyodesenteria* is the agent of the swine dysentery, *B. intermedia* is the pathogen mainly of the adult chickens and *B. Pilosicoli* is the one which is responsible for the cause of intestinal *spirochetosis*. This *B. Pilosicoli* has a wide range of infection when compared to the other two main pathogenic species which have been colonizing in the mammals and birds as well as in human beings [Trott DJ et al 1998]. The colonizing of the *B. Pilosicoli* species has been rapidly grown in developing countries due to unhygienic conditions. The infected pigs and chickens in the farm show the depressed rate of growth and production [Trott DJ et al 1997, Margawani KR, et al 2004, Munshi MA et al 2004, Nelson EJ et al 2009].

Rectal bleeding, chronic diarrhea with abdominal pain and focal colitis have been observed in the individuals who are suffering from the B. Pilosicoli infection. In an in vitro study, it showed that by using Caco-2 cells has shown that B. Pilosicoli strains initially targets the cell junctions where one cell end of the spirochete invaginates into the Caco-2 cell membranes [Naresh R, et al 2009]. When the mode of infection has been started here the whole cell surface will be colonized by the B. Pilosicoli species. Then a false brush border will be formed by the infections In this model colonized monolayers demonstrated accumulation of the actin at the cell junction, loss of tight junction integrity, fragmentation and condensation of the nuclear material consistent with apoptosis, here a spirochaetamia with B. Pilosicoli has been recorded in the immunocompromised human beings [ Trott DJ et al 1997, Bait-Merabet L et al 2008] Brachyspira Pilosicoli is a food born and water-born disease i;e the water and food sources which are contaminated by the B.Pilosicoli [Margawani KR et al 2004, Oxberry SL et al 1998, Verlinden M, et al 2012]. Of these two species i.e; B. Hydosynteria and B.Pilosicoli here B. Pilosicoli has been shown that about 30% of the individuals have been colonized by this species including Oman [Barrett SP et al 1990] and Papua New Guinea [Trott DJ 1997], but recent investigations showed that it is uncommon in developed countries [Lee JI et al 1992, Tompkins DS et al 1986, Brooke CJ et al 2001] No epidemiological studies have been conducted on the B. Aalborg because this organism is the one which is fastidious and taking up to three weeks to grow under anaerobic incubation on the specialized media. Till today B. Aalborg have been found in the humans and here it has been isolated in the five studies i;e in three it was grown in the colorectal biopsies [Hovind-Hougen K et al 1981, Kraaz W, et al 2000] and in two it was grown from feces [Brooke CJ et al 2003, Calderaro A et al 2003].

Table 1: Brachyspira species, their host, and pathogenicity

Species	Host	Pathogenicity
B. aalborgi	Human, primates	Mild to moderate
	Human, primates	(Hovind-Hougen K et al 1981)
B. alvinipulli	Chicken, goose, Red-breasted, merganser,	Mild to severe
	dog	(Stanton TB et al 2015)
B. hyodysenteriae	Chicken, goose, mallard, common rhea, pig,	Severe (Harris DL et al 1972,
	rat, mouse	Taylor DJ et al 1971)
B. innocent	Chicken, pig, dog, horse	None (Kinyon JM et al 1979,
		Stanton TB. et al 1992)
B. intermedia	Chieken Die	Mild to moderate
	Chicken, Pig	(Stanton TB et al 1997)
B. murdochii	Chicken, pig, rat	None (Stanton TB et al 1997)
B. pilosicoli	Chicken, pheasant, grey partridge, feral	Mild to moderate
	water birds, common rhea, pig, dog, human.	(Trott DJ, et al 1996)

#### 2. OVERVIEW OF THE DISEASE

#### 2.1 Signs and symptoms

The infections of the *Brachyspira Pilosicoli* are observed commonly in the egg-laying chickens in the poultry farms. Many cases have been reported worldwide especially in Europe, the US and Australian countries where a huge number of chickens have been reared, so here the Gastro-Intestinal infections have been observed commonly here [Hampson DJ, et al 2013, Stephens CP et al 1999]. Infection of the *Brachyspira Pilosicoli* starts from milder to higher, which leads to higher death rate of chickens[Hampson DJ, et al 2013, Stephens CP, et al 1999, Erlandson K et al 2005] Before, at the initial stage of infection the diarrhea, feces with altered color and consistency will be observed [Taylor P et al 1993]. This infection starts resulting in the low growth rate of the chickens and layers will be delayed up to 7-9 weeks in the egg-laying processes and also the poor egg quality will be produced by the infected hens. [Taylor P et al 1993, Griffiths IB, et al 1987, Taylor P et al 2002].

The laid egg will be sometimes cracked and the outer shell will be contaminated by feces [Swayne DE et al 1992]. The chicks hatched by the infected mother will have the same similarity i.e.; decreased weight paler feces, delayed lay one set, despite the absence of the contamination. These results made raised questions of the potential epigenetic variations by infections to *Brachyspira Pilosicoli*. At the intestinal level, the cross-section of the infected chickens shows that the *Brachyspira pilosicoli* fixed to the cells of the intestinal wall. The attached part's tissues look often like bleeding, the intestinal wall shows the loss of microvilli. Here due to the loss of microvilli the increased amount of water will be seen in feces [Taylor P et al 1993, Beardsworth PM et al 2004].

## 2.2 Zoonotic potential

In humans it is mostly seen in the immune-compromised patients, this is confirmed by biopsy i;e it shows its symptoms through diarrhea and through abdominal pain [Trott DJ et al 1995, Calderaro A et al 2007]. As a result of septicemia in some of the rare cases, this infection causes death in human beings [Prim N, et al 2011]. These cases have been only observed in the elderly and immunocompromised patients or in the population having a dense area with poor hygienic conditions [Lee JI et al 1992, Trott DJ et al 1997, Margawani KR et al 2004]. It has been notified that the *B. Pilosicoli* is able to survive and be transmitted to the consumers via contaminated meat of chickens and pigs which has been infected [Verlinden M, et al 2012].

In 2012, Mappley et al. [Mappley LJ, et al 2012] carried out the genetic comparison of the three strains of the *B. Pilosicoli* which was isolated from the humans, pigs, and chickens, which showed the genotype of these three strains were very similar. But some difference was notified in the genome size and arrangement and in some putative coding regions for the carbohydrates, amino acids and nucleotide metabolism transport [Mappley LJ, et al 2012]. These data highlight some fundamental genetic differences between these species isolated from the chicken and humans in comparison to strains isolated from pigs. [Mappley LJ, et al 2012].

## 3. MATERIALS AND METHODS

**EDGAR** ("Efficient Database framework for comparative Genome Analyses using BLAST score Ratios") an online available search tool helped in taking the *B. pilosicoli* bacteria for the comparative analysis i;e to compute the core genome, the pan-genome, singleton genes, synteny plots, and Venn diagrams. Here the species of the *Brachyspira Pilosicoli* were selected in the parameter selection. A minimum of five species of *B. Pilosicoli* was selected in the parameter selection. Venn diagram was built in the EDGAR which built the core genome contained about 1378 protein sequences. The availability of these protein sequences was found in EDGAR bioinformatics and they were downloaded and converted to FASTA format. FASTA format protein sequences were then blasted using the NCBI BLAST tool which finds regions of similarity between biological sequences. The program compares protein sequences to sequence databases and calculates the statistical significance. Next, the studies say that the mortality rate and the number of infected organisms were the humans, hens, and pigs, in these species the bacteria colonized in the intestinal parts of it and showed its pathogenicity. The availability of the genome sequence was seen in the NCBI BLAST and it was confirmed that the availability was there.

Next, in the blast, the protein query was entered individually than in choosing the search set options the reference protein (Refseq\_protein) was selected. While choosing the organisms here pigs (taxid: 9821), pigs < #2&gt; (taxid: 9823) and chickens (taxid: 9031) were selected. These mentioned organisms were selected because of the pathogenesis of the bacteria in these species was very high. Then in the program selection here, DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST) Algorithm was selected. Next, the whole built options were BLAST then here. The Whole 1378 proteins were BLAST using the

Protein Blast here and the result obtained was further noted, it had 28 proteins sequences had no similarities out of 1378 protein sequences. Here we considered the no similarities found sequences as the drug targets which were then submitted to the DEG BLAST. The submitted protein sequences were then DEG BLAST here and the result obtained was here out of 28 query sequences 18 of them were found to be no similar against the Prokaryotes (no hit), single gene and multiple genes were also considered and here 18 query sequence were taken out, 9 of them were left out and CEG (Cluster Of Essential genes) was done. Results obtained showed us that further studies can be carried into consideration with the protein models which can be built using obtained protein sequences.

#### 4. RESULT

Table 2: Protein query and their functions

Protein Query	Protein name	Biological Process	Role in Virulence
BP951000 0622	MerR family transcriptional regulator	GO:0006355 regulation of transcription, DNA-templated	Metal-binding domain (Brown, N. L., et al 2003, Changela, A., et al 2003, Matthew J. Bush et al 2017)
BP951000_0591	methionine-tRNA ligase	None predicted	Catalyzes the chemical reaction (Leung K-Y et al 2013, Van Haandel L et al et al 2012, Tian Q et al 2015& Kozo Tomita 2017)
BP951000_0785	methyl-accepting chemotaxis protein B	GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000_1875	methyl-accepting chemotaxis sensory transducer	GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000_1776	methyl-accepting chemotaxis protein B	GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000_1844	methyl-accepting chemotaxis sensory transducer	GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000_1845	methyl-accepting chemotaxis sensory transducer	GO:0006935 chemotaxis GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000 0500	methyl-accepting chemotaxis protein B	GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000_1276	methyl-accepting chemotaxis protein B	GO:0006935 chemotaxis GO:0007165 signal transduction	Sensing environmental cues (Abu Iftiaf Md Salah Ud-Din1 et al 2017, Berleman JE et al 2005 & He K, Bauer CE (2014))
BP951000 0882	methionyl-tRNA formyltransferase	GO:0009058 biosynthetic process GO:0071951 conversion of methionyl-tRNA to N-formyl-methionyl-tRNA	Involvement in the biosynthetic process (Xi Peng et al 2016 & Zhang, Y. M., J. K. Liu and T. Y. Wong (2003))
BP951000_2210	metal-dependent amidohydrolase	None predicted	Helps in enzyme Biodegradation (Alexandre A. de Castro 2016, Singh BK (2009), Theriot CM, Grunden AM (2011) & Bigley AN, Raushel FM (2013))
BP951000_1654	methyltransferase	None predicted.	Epigenetic modification (Siyi Chena, et al 2017, W. Reik, J. Walter 2001 & K.D. Robertson 2005)
BP951000 0520	membrane-bound lytic murein transglycosylase D	GO:0000270 peptidoglycans metabolic process	The essential catalyst of bacterial cell wall function (E. Scheurwater, 2008, Dusan Hesek, et al 2017)
BP951000_0389	metal-dependent phosphohydrolase	None predicted.	regulation of response against nutritional stress conditions (Shama Khan1 et al 2015, Gross M, Marianovsky I, Glaser G (2006))

BP951000_1097	methyltransferase GidB	GO:0006364 rRNA processing	Epigenetic modification (Siyi Chena et al 2017, W. Reik et al 2001 & K.D. Robertsn et al 2005)
BP951000 1849	metal-dependent amidase aminoacylase carboxypeptidase	GO:0008152 metabolic process	Involvement in Peptidolytic growth (Veronica M., Lin, L et al 2007)
BP951000 0058	metal-dependent amidase/aminoacylas e/carboxypeptidase AbgB	GO:0008152 metabolic process	Involvement in Peptidolytic growth (Lin, L et al 2007)
BP951000 0378	methylenetetrahydrof olate reductase	GO:0006555 methionine metabolic process GO:0055114 oxidation-reduction process	Causes gene mutation in chromosome 1 (Mark W. Morningstar, et al 2017)

## 5. DISCUSSION

The above-mentioned proteins are the target proteins and they were obtained because of their high pathogenicity factor and involvement in the metabolic activities, mutation, hydrolase activity, signal transduction and many more.

In our subtractive genomics approach, the essential genome sequences were available in NCBI Protein BLAST, in which no similarities were found in only between 28 proteins sequences virulence caused ones were brought back from the NCBI BLAST and searched against Database of Essential Genes (DEG) tool respectively. When searched under DEG BLAST showed only 18 of them showed the no similarities (no hit) which were considered as the druggable targets. The Interpro an online tool helped to known the individual proteins' biological process and molecular function and the research studies on these proteins showed us that the involvement of them in the virulence factors and showed the matching of them with the other proteins. Hence concluded that the Bacteria *B. Pilosicoli* selected these above-mentioned proteins as its targets because of their involvement in the virulence factors, which make them differ from other bacteria.

#### 6. CONCLUSION

With consideration to the above results, it can be concluded that the obtained proteomes have the highest rate of pathogenicity role in hens, pigs and also zoonotic potentials. There is a need for investigations regarding this proteomics which play a major role with respect to their pathogenicity. Further studies can be carried out by using these proteomes to feature investigate the protein models to be built and to find out the drug targets to the above-obtained results. The intention of ligand-protein docking is to expect the most important binding mode(s) of a ligand with a protein of recognized three-dimensional structure. Successful docking techniques seek high-dimensional spaces correctly and use a scoring feature that efficaciously ranks candidate dockings, they also can be used to carry out virtual screening on large libraries of compounds, rank the results, and recommend structural hypotheses of the way the ligands inhibit the target, that's useful in lead optimization.

## 7. REFERENCES

- [1] Abu Iftiaf Md Salah Ud-Din, Anna Roujeini kova1, Methyl-accepting chemotaxis proteins: a core sensing element in prokaryotes and archaea © Springer International Publishing 2017.
- [2] Alexandre A. de Castro 1, Letícia C. Assis 1, Daniela R. Silva 1, Silviana Corrêa 1, Tamiris M. Assis 1, Giovanna C. Gajo 1, Flávia V. Soares 1 and Teodorico C. Ramalho 1,2,\* Computational enzymology for degradation of chemical warfare agents: promising technologies for remediation processes 1 Department of Chemistry, Federal University of Lavras, 2016.
- [3] Bait-Merabet L, Thille A, Legrand P, Brun-Buisson C, Cattoir V: Brachyspira pilosicoli bloodstream infections: case report and review of the literature. Ann Clin Microbiol Antimicrob 2008, 7-19.
- [4] Barrett SP. Intestinal spirochaetes in a Gulf Arab population. Epidemiol Infect 1990; 104: 261-266.
- [5] Beardsworth PM, Hernandez JM. Yolk color an important egg quality attribute. Int Poult Prod 2004; 12: 17-8
- [6] Berleman JE, Bauer CE (2005) Involvement of a Che-like signal transduction cascade in regulating cyst cell development in Rhodospirillum centum. Mol Microbiol 56:1457–1466.
- [7] Bigley AN, Raushel FM (2013) Catalytic mechanisms for phosphotriesterases. Biochim Biophys Acta 1834: 443–453.
- [8] Brooke CJ, Clair AN, Mikosza ASJ, Riley TV, Hampson DJ. Carriage of intestinal spirochaetes by humans: epidemiological data from Western Australia. Epidemiol Infect 2001; 127-369
- [9] Brooke CJ, Riley TV, Hampson DJ. Evaluation of selective media for the isolation of Brachyspira and borgi from human feces. J Med Microbiol 2003; 52: 509-513.
- [10] Brown, N. L., Stoyanov, J. V., Kidd, S. P., and Hobman, J. L. (2003). The MerR family of transcriptional regulators. FEMS Microbiol. Rev. 27, 145–163.
- [11] Calderaro A, Gorrini C, Peruzzi S, Piccolo G, Dettori G, Chezzi C. Occurrence of human intestinal spirochetosis in comparison with infections by other enteropathogenic agents in an area of Northern Italy. Diagn Microbiol Infect Dis 2007; 59: 157-63.
- [12] Calderaro A, Villanacci V, Conte M, et al. Rapid detection and identification of Brachyspira Aalborg from rectal biopsies and feces of a patient. Res Micro Biol 2003; 154: 145-153
- [13] Changela, A., Chen, K., Xue, Y., Holschen, J., Outten, C. E., O'Halloran, T. V., et al. (2003). Molecular basis of metal-ion selectivity and femtomolar sensitivity by CueR. Science 301, 1383–1387.
- [14] Dusan Hasek, David A. Dik, Jennifer Fishovitz, Elena Lastochkin, Bill Boggess, JedF.Fisher, and Shahriar Mobashery\* From Genome to Proteome to Elucidation of Reactions for All Eleven Known Lytic Transglycosylases from Pseudomonas aeruginosa Mijoon Lee 2017.

- [15] E. Scheurwater, C. W. Reid, A. J. Clarke, Int. J. Biochem. Cell Biol. 2008.
- [16] Erlandson K, Klinger E. Intestinal spirochaetosis: epidemiology, microbiology, and clinical significance. Clin Microbiol Newslett 2005; 27: 91-6.
- [17] Griffiths IB, Hunt BW, Lister SA, Lamont MH. Retarded growth rate and delayed onset of egg production associated with spirochaete infection in pullets. Vet Rec 1987; 121: 35-7.
- [18] Gross M, Marianovsky I, and Glaser G (2006) MazG–a regulator of programmed cell death in Escherichia coli. Mol Microbiol 59:590–601
- [19] Hampson DJ, Swayne DE, Glisson JR, McDougald LR, Nolan LK, Suarez DL. The disease of poultry. Ames: Blackwell; 2013.
- [20] Hampson, D. J., Oxberry, S. L. & La, T. (2006). Potential for zoonotic transmission of Brachyspira pilosicoli. Emerg Infect Dis 12, 869–870.
- [21] Harris DL, Glock RD, Christensen CR, Kinyon JM. Inoculation of pigs with Treponema hyodysenteriae (new species) and reproduction of the disease. Vet Med Small Anim Clin 1972; 67: 614.
- [22] He K, Bauer CE (2014) chemosensory signaling systems that control bacterial survival. Trends Microbiol 22:389–398.
- [23] Hovind-Hougen, K., Ellis, W. A. & Birch-Andersen, A. (1981). Leptospira parva sp. nov.: some morphological and biological characters. Zentralbl Bakteriol Mikrobiol Hyg A 250, 343–354.
- [24] K.D. Robertson, DNA methylation and human disease, Nat. Rev. Genet., 6 (2005),597–610.
- [25] Kinyon JM, Harris DL. Treponema innocent, a new species of intestinal bacteria, and emended description of the type strain of Treponema hyodysenteriae. Int J Syst Bacteriol 1979; 29: 1029.
- [26] Kozo Tomita E-mail: kozo\_tomita@cbms.k.u-tokyo.ac.jp. A possible link between specific transfer RNA methylation and tumorigenic phenotype of breast cancer, 2017.
- [27] Kraaz W, Pettersson B, Thunberg U, Engstrand L, Fellstrom C. Brachyspira Aalborg infection diagnosed by culture and 16S ribosomal DNA sequencing using human colonic biopsy specimens. J Clin Microbiol 2000; 39: 3555-3560.
- [28] Lee JI, Hampson DJ. Intestinal spirochaetes colonizing Aborigines from communities in the remote north of Western Australia. Epidemiol Infect 1992; 109:133-141.
- [29] Leung K-Y et al. Folate metabolite profiling in different cell types and embryos suggests variation in folate one-carbon metabolism, including developmental changes in the human embryonic brain. Mol Cell Biochem 2013; 378: 229-236.
- [30] Lin, L. L., Chen, M. H., Chien, H. C., Kan, S. C. et al., Characterization of a bifunctional amino acylase/carboxypeptidase from radioresistant bacterium Deinococcus radiodurans R1. J. Biotechnol. 2007, 128, 322–334.
- [31] Mappley LJ, Black ML, Abuoun M, Darby AC, Woodward MJ, Parkhill J, et al. Comparative genomics of Brachyspira pilosicoli strains: genome rearrangements, reductions, and correlation of genetic complement with phenotypic diversity. BMC Genomics 2012; 13: 454.
- [32] Margawani KR, Robertson ID, Brooke CJ, Hampson DJ: Prevalence, risk factors and molecular epidemiology of Brachyspira pilosicoli in humans on the island of Bali, Indonesia. J Med Microbiol 2004, 53:325–332.
- [33] Mark W. Morningstar 1\*, Megan N. Strauchman 1, Clayton J. Stitzel 2, Brian Dovorany 3, Aatif Siddiqui 4. Methylenetetra hydro folate Reductase (MTHFR) Gene Mutations in Patients with Idiopathic Scoliosis: A Clinical Chart Review 1 Natural Wellness & Pain Relief Center, Grand Blanc, MI, USA 2 Lancaster Spinal Health Center, Lititz, PA, USA 3 Posture & Spine Care Center, Green Bay, WI, USA 4 Esprit Wellness, New York, NY, USA, 2017.
- [34] Matthew J. Bush, Govind Chandra, 1 Kim C. Findlay and Mark J. Buttner Department of Molecular Microbiology, John Innes Centre, Norwich Research Park, Norwich NR4 7UH, UK. Department of Cell and Developmental Biology, John Innes Centre, Norwich Research Park, 2017.
- [35] Munshi MA, Traub RJ, Robertson ID, Mikosza ASJ, Hampson DJ: Colonization and risk factors for Brachyspira Aalborg and Brachyspira pilosicoli in humans and dogs on tea-estates in Assam, India. Epidemiol Infect 2004, 132:137–144.
- [36] Naresh R, Song Y, Hampson DJ: The intestinal spirochete Brachyspira pilosicoli attaches to cultured Caco-2 cells and induces pathological changes. PLoS ONE 2009, 4(12).
- [37] Nelson EJ, Tanudra A, Chowdhury A, Kane AV, Qadri F, Calderwood SB, Coburn J, Camilli A: High prevalence of spirochetosis in cholera patients, Bangladesh. Emerg Infect Dis 2009, 15:571–573.
- [38] Oxberry SL, Trott DJ, Hampson DJ: Serpulina pilosicoli, water birds and water: potential sources of infection for humans and other animals. Epidemiol Infect 1998, 121:219–225.
- [39] Prim N, Pericas R, Espana ol M, Rivera A, Mirelis B, Coll P. Bloodstream infection due to Brachyspira pilosicoli in a patient with multiorgan failure. J Clin Microbiol 2011; 49: 3697-9.
- [40] Shama Khan1 Mohd. Shahbaaz2 Krishna Bisetty2 Faizan Ahmad1 Md. Imtiaz Hassan1 Classification and Functional Analyses of Putative Conserved Proteins from Chlamydophila pneumoniae CWL029. International Association of Scientists in the Interdisciplinary Areas and Springer-Verlag Berlin Heidelberg 2015.
- [41] Singh BK (2009) Organophosphorus-degrading bacteria: ecology and industrial applications. Nat Rev Microbiol 7: 156–164.
- [42] Siyi Chena, Huan Maa, Wang Lia, b, Zhou Niea, Shouzhuo Yaoa. 2017 An entropy-driven signal amplifying strategy for real-time monitoring of DNA methylation process and high-throughput screening of methyltransferase inhibitors, 2017.
- [43] Stanton TB, Hampson DJ. Physiology of ruminal and intestinal spirochaetes. Madison, USA: CAB International; 1997.
- [44] Stanton TB, Lebo DF. Treponema hyodysenteriae growth under various culture conditions. Vet Microbiol 2015; 18: 17790
- [45] Stanton TB. Proposal to change the genus designation Serpula to Serpulina containing the species Serpulina hyodysenteriae and Serpulina innocent. Int J Syst Bacteriol 1992; 42: 89-90.
- [46] Stanton TB: The genus Brachyspira. In The Prokaryotes. Volume 7. Edited by Falkow S, Rosenberg SE, Schleifer KH, Stackebrant E. New York: Springer; 2006:330–356.
- [47] Stephens CP, Hampson DJ. Prevalence and disease association of intestinal spirochaetes in chickens in Eastern Australia. Avian Pathol 1999; 28: 447-454.
- [48] Swayne DE, Bermudez AJ, Saqartz KA, Monfort JD, Stoutenburg JW, Hayes JR. Association of cecal spirochetes with pasty vents and dirty eggshells in layers. Avian Dis 1992; 36:76-81.

- [49] Taylor DJ, Alexander TJ. The production of dysentery in swine by feeding cultures containing a spirochaete. Br Vet J 1971; 127: 58-61.
- [50] Taylor P, Dwars RM, Davelaar FG, Smit HF. Infection of broiler parent hens with avian intestinal spirochaetes: effects on egg production and chick quality. Avian Pathol 1993; 22: 37-41.
- [51] Taylor P, Stephens CP, Hampson DJ. Experimental infection of broiler breeder hens with the intestinal spirochaete Brachyspira (Serpulina) pilosicoli causes reduced egg production. Avian Pathol 2002; 31: 37-41.
- [52] Theriot CM, Grunden AM (2011) Hydrolysis of organophosphorus compounds by microbial enzymes. Appl Microbiol Biotechnol 89: 35–43.
- [53] Tian Q, Wang C, Liu Y, Xie W. Structural basis for recognition of G-1-containing tRNA by histidyl-tRNA synthetase. Nucleic Acids Res 2015; 43: 2980-2990.
- [54] Tompkins DS, Foulkes SJ, Godwin PGR, West AP. Isolation and characterization of intestinal spirochaetes. J Clin Pathol 1986; 39: 535-541.
- [55] Trott DJ, Combs BG, Mikosza AS, Oxberry SL, Robertson ID, Passey M, et al. The prevalence of Serpulina pilosicoli in humans and domestic animals in the Eastern Highlands of Papua New Guinea. Epidemiol Infect 1997; 119: 369-379.
- [56] Trott DJ, McLaren AJ, Hampson DJ. Pathogenicity of human and porcine intestinal spirochetes in one-day-old specific pathogen-free chicks: an animal model of intestinal spirochetosis. Infect Immun 1995; 63: 05-10.
- [57] Trott DJ, Mikosza ASJ, Combs BG, Oxberry SL, Hampson DJ: Population genetic analysis of Serpulina pilosicoli and its molecular epidemiology in villages in the Eastern Highlands of Papua New Guinea. Int J Syst Bacteriol 1998, 48:659–668.
- [58] Trott DJ, Oxberry SL, Hampson DJ: Evidence for Serpulina hyodysenteriae being recombinant, with an epidemic population structure. Microbiology 1997, 143:3357–3365.
- [59] Trott DJ, Stanton TB, Jensen NS, Duhamel GE, Johnson JL, Hampson DJ. Serpulina pilosicoli the agent of porcine intestinal spirochetosis. Int J Syst Bacteriol 1996; 46:15-20.
- [60] Van Haandel L et al. Comprehensive quantitative measurement of folate polyglutamate in human erythrocytes by ion-pairing ultra-performance liquid chromatography/tandem mass spectrometry. Rapid Commun Mass Spectrom 2012; 26: 1617-1630.
- [61] Verlinden M, Pasmans F, Garmyn A, De Zutter L, Haesebrouck F, Martel A: Occurrence of viable Brachyspira spp. on carcasses of spent laying hens from supermarkets. Food Microbiol 2012, 32:321–324.
- [62] Veronica M. Jarocki1, Jessica L. Tacchi1, and Steven P. Djordjevic1, Non-proteolytic functions of microbial proteases increase pathological complexity, 2007.
- [63] W. Reik, J. Walter Genomic imprinting: parental influence on the genome Nat. Rev. Genet., 2 (2001), 21–32
- [64] Xi Peng1†, Jie Yang2†, and Yang Gao1\*Proteomic Analyses of Changes in Synechococcus sp. PCC7942 Following UV-C Stress. 2017.
- [65] Zhang, Y. M., J. K. Liu, and T. Y. Wong (2003) The DNA excision repair system of the highly radioresistant bacterium Deinococcus radiodurans is facilitated by the pentose phosphate pathway. Mol. Microbiol. 48, 1317–1323.